# Flunisolide – Triamcinolone acetonide. MS differentiation in positive and in negative ionization mode. LC differentiation.

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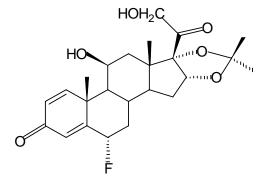
### Introduction

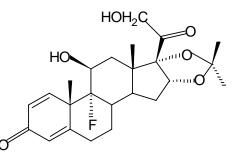
Flunisolide and Triamcinolone acetonide belong to the glucocorticosteroid class. All glucocorticosteroids are prohibited in competition and included in the WADA Prohibited List in the section S9 [1].

The two glucocorticosteroids have the same molecular mass (434), same molecular formula  $(C_{24}H_{31}FO_6)$  and similar chemical structures; they differ only by the position of the fluorine atom on the steroid skeleton –  $6\alpha$  for Flunisolide, respectively,  $9\alpha$  for Triamcinolone acetonide.

Therefore, the two prohibited substances may have common ions or MRM transitions and close chromatographic retention times, therefore interfering in each others detection. In the case of the analyze of a doping control sample containing one of these two substances, these interferences may result in a false negative analytical finding or in a misidentification of the substance detected.

This paper presents the results of the tests carried-out by the liquid chromatography coupled with tandem mass spectrometry with triple quadrupole in order to differentiate the two prohibited substances.





Flunisolide  $C_{24}H_{31}FO_6$  M=434

Triamcinolone acetonide  $C_{24}H_{31}FO_6$  M=434

Figure 1. Chemical structure of Flunisolide and Triamcinolone acetonide

#### Materials and Methods

The tests have been carried-out on  $10\mu$ g/ml solutions in methanol of reference materials. The instrumental analysis has been carried-out on two analytic equipments: AGILENT 6410, respectively, VARIAN 1200L. The analytic conditions are presented in table 1.

		A
Equipment	AGILENT 6410	VARIAN 1200L
Column	ZORBAX SB-C18	ChromSep SS OmniSpher 3 C18
	(50x2.1mm, 5µm)	(100x2.0mm, 3µm)
Column Thermostat	$30^{0}$ C	25°C
Solvent A	5mM NH <sub>4</sub> HCOO in water	0.1% acetic acid and 5mM
		$NH_4CH_3COO$ in water (v/v)
Solvent B	5mM NH <sub>4</sub> HCOO in acetonitrile	Methanol
Flow	0.3ml/min	0.25ml/min
Gradient B	30-50% in 1min, 50%-70% in	30-50% in 1min, 50%-70% in
	3min, 5min at 70%, 5min at 30%	3min, 1min at 70%, 5min at 30%
Injection volume	2μL	10µL
Ionization	ESI positive or negative:	APCI negative:
	Drying gas: $121 \text{ N}_2/\text{min}$ at $350^{\circ}\text{C}$	Drying gas: 12psi N <sub>2</sub> at 150 <sup>o</sup> C
	Nebulysing gas: 50psi N <sub>2</sub>	Nebulysing gas: 58psi air
		Auxiliary gas: 17psi N <sub>2</sub> at $400^{\circ}$ C
	Capillary needle: 4000V	Corona current: 5µA
Collision gas	Nitrogen	Argon, 1.5mTorr
MRM transitions	Analysis in Product Ion Scan	Use of the MSMS Breakdown
selection and	mode followed by analyses in	acquisition soft during direct
collision energy	MRM mode at various collision	injection of a 10µg/ml solution of
optimization	energies	reference material

Table 1. LC/MS <sup>2</sup> anal	vtic parameters
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### Results and Discussion

### MS Differentiation

Both substances (like the majority of the glucocorticosteroids) give good signals both in ESI and in APCI and ionize both in positive mode, forming  $[M+H]^+$ , and in negative mode, forming adducts with anions from the mobile phase [2,3,4,5].

In figure 2 the Product Ion spectra, at 10V collision energy, in positive mode, are shown for Flunisolide (a) and Triamcinolone acetonide (b) respectively. In table 2, the selected MRM transitions together with the optimized collision energy are given. Table 2 also allows comparison of the (relative) abundances of these selected transitions between both compounds. The ion 417 (loss of H<sub>2</sub>O) is specific to Flunisolide, while the ions 415 (loss of HF) and 357 to Triamcinolone acetonide, but even for the common ions, 339 and 321, the relative abundances are too different to satisfy the WADA identification criteria [6] concerning the mass spectrometry if one would try to confirm a sample containing Flunisolide against a reference with Triamcinolone acetonide (or vice versa).

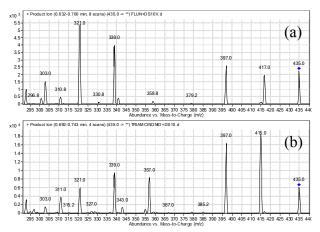


Figure 2. Product Ions Scans in positive ionization mode (Agilent 6410)

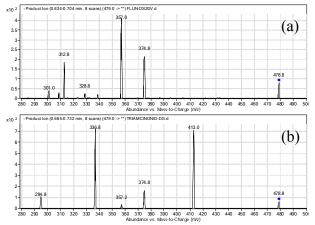


Figure 3. Product Ions Scans in negative ionization mode (Agilent 6410, formate adduct as precursor ion)

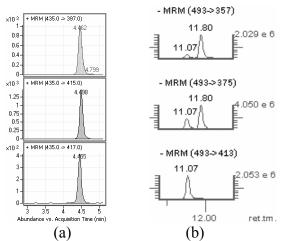


Figure 4. LC/MS<sup>2</sup> analysis of urines spiked with 20ng/ml Flunisolide and Triamcinolone acetonide, prepared and analyzed by the routine screening procedure [5]: (a) on Agilent 6410 in positive mode with water/ acetonitrile, (b) on Varian 1200L in negative mode (acetate adducts) with water/methanol

Table 2. Optimized collision energy MRM's abundances on Agilent 6410 in positive ionization mode

MRM	Flunisolide		Triamcinolone	
(collision energy)	(Area)		acetonide (Area)	
(+) 435->321 (10∨)	74532	159%	56012	50%
(+) 435->339 (5∨)	61752	132%	77323	69%
(+) 435->357 (5∨)	684	1%	67586	60%
(+) 435->397 (5∨)	46767	100%	112325	100%
(+) 435->415 ( 5∨)	3443	7%	218793	195%
(+) 435->417 (5∨)	38770	83%	597	1%

Table 3. Optimized collision energy MRM's abundances on Agilent 6410 in negative ionization mode (formate adduct as precursor ion)

MRM	Flunisolide		Triamcinolone	
(collision energy)	(Area)		acetonide (Area)	
(-) 479->313 (30∨)	1329	12%	-	-
(-) 479->337 (30∨)	396	4%	4825	70%
(-) 479->357 (15∨)	6895	63%	1069	16%
(-) 479->375 (15∨)	10881	100%	6853	100%
(-) 479->413 (15∨)	-	-	8036	117%

Table 4. Optimized collision energy MRM's abundances on Varian 1200L in negative ionization mode (acetate adduct as precursor ion)

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MRM	Flunisolide		Triamcinolone	
(collision energy)	(h.10	<sup>-3</sup> )	acetonide	(h.10 <sup>-3</sup> )
(-) 493->185 (28∨)	10990	10%	-	-
(-) 493->357 (21∨)	50350	44%	8010	19%
(-) 493->375 (14∨)	114200	100%	42700	100%
(-) 493->413 (22∨)	-	-	50330	118%

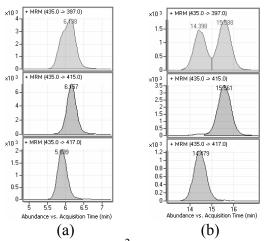


Figure 5.  $LC/MS^2$  analysis of a 5µg/ml Flunisolide and Triamcinolone acetonide solution in methanol analyzed on Agilent 6410 in positive mode with water/acetonitrile on a C18 column eluted: (a) isocratic with 30% solvent B, (b) isocratic with 25% solvent B

In figure 3 (a) and (b) and in table 3 the Product Ion spectra are shown, at 20V collision energy, in negative mode (formate adduct, 479, as precursor ion) and the MRM abundances at optimized collision energy, obtained on Agilent 6410 for the two compounds. The ion 313 is specific to Flunisolide, while 413 and 337 to Triamcinolone acetonide

In table 4 the MRM abundances are shown, at optimized collision energy, obtained in negative mode (acetate adduct, 493, as precursor ion) on Varian 1200L for the two compounds. The ion 185 is specific to Flunisolide, while 413 to Triamcinolone acetonide.

## LC Differentiation

In the routine screening analysis chromatographic conditions, the Flunisolide – Triamcinolone acetonide chromatographic separation succeeds when using, on Varian 1200L, water/methanol based mobile phase (figure 4b), but not when using, on Agilent 6410, water/acetonitrile based mobile phase (figure 4a).

In isocratic conditions with 30% solvent acetonitrile (figure 5a), although the peaks are not at all separated, the difference between retention times (0.258min, 4.2%) >4%, which is already sufficient to differentiate: if one would try to confirm a sample containing Flunisolide against a reference with Triamcinolone acetonide (or vice versa), the WADA identification criteria [6] concerning the liquid chromatography (retention time  $\pm$  2%) would not be satisfied.

In isocratic conditions with 25% solvent B acetonitrile (figure 5b) the chromatographic separation looks sufficient to avoid major interferences between the two compounds.

### References

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